Samarium Iodide-Mediated Reformatsky Reactions for the Stereoselective Preparation of β -Hydroxy- γ -amino Acids: Synthesis of Isostatine and Dolaisoleucine

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S Supporting Information

ABSTRACT: The synthesis of β -hydroxy-y-amino acids via SmI₂-mediated Reformatsky reactions of α -chloroacetyloxazolidinones with aminoaldehydes is reported. Diastereoselective coupling is demonstrated to depend on the absolute configuration of the Evans chiral auxiliary employed in the reaction, allowing erythro or threo products to be obtained selectively. The potential utility of the methodology is exemplified by the facile synthesis of biologically relevant N-Boc-isostatine (2b) and N-Bocdolaisoleucine (3c).

Nonproteinogenic amino acids containing ^β-hydroxy-γamino acid motifs comprise a biologically important class of agents that includes (3S,4S)-4-amino-3-hydroxy-6-methylheptanoic acid (statine, 1), a component of aspartic protease inhibitors such as peptstatine¹ and $(3S, 4R, 5S)$ -4-amino-3hydroxy-5-methylheptanoic acid (isostatine, 2a), found in the cytotoxic didemnin cyclodepsi[pe](#page-4-0)ptides.² Additional examples are (3S,4R,5S)-3-methoxy-5-methyl-4-(methylamino)heptanoic acid (dolaisoleucine or "Dil", 3a) and [\(](#page-4-0)2R,3R)-3-methoxy-2 methyl-3-((S)-pyrrolidin-2-yl)propanoic acid (dolaproine or "Dap", 4), both of which are key constituents of cytotoxic peptides, including dolastatin $10³$ as well as $(2S, 3S, 4R)$ -4amino-3-hydroxy-2-methylpentanoic acid (5), a component of the antin[eo](#page-4-0)plastic glycopeptide, bleomycin B2 (Figure 1).⁴

Several synthetic methodologies have been reported for the stereoselective syntheses of β -hydroxy- γ -amino acids (reviewed

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in ref 5). Of particular note for the preparation of analogues such as 4^6 and 5^7 that contain substituents at the α -position are "doub[le](#page-4-0) stereo-differentiating" ⁸ aldol reactions of amino aldehyde[s](#page-4-0) with [c](#page-4-0)hiral acyloxazolidinones, where nascent α functionality within the auxilia[ry](#page-4-0) facilitates chirality transfer. As shown in Scheme 1 (eq 1), the aldol addition of propanoyloxazolidinone 7a to N-Boc- (S) -prolinal 6 provides the Dap precursor 8 as a s[in](#page-1-0)gle diastereomer. In contrast, similar chemistry applied to the preparation of products that lack α substituents can result in significantly less effective stereoinduction. This is exemplified by the synthesis of N-Cbz Dil $(3b)$ through the reaction of N-Cbz-N-methyl (S) -isoleucinal (9) with the boron enolate of acetyloxazolidinone 7b, where a near complete lack of stereocontrol is observed at the βhydroxyl center of the resulting product $10a$ (Scheme 1, eq 2).⁹ In this latter example, limitations imposed by the absence of an α α α -[s](#page-4-0)ubstitutent can be overcome through a two-step process involving the aldol reaction of a methylthio-containing acyloxazolidinone (7c), which yields a thiomethyl group at the α -position (10b) that facilitates chirality transfer to the β hydroxyl center. Once the stereodirecting role of the thiomethyl group has been achieved, it can be removed reductively to provide the desired final product, N-Cbz Dil $(3b).⁹$

Metal-catalyzed Reformatsky reactions between aldehydes or keto[ne](#page-4-0)s and compounds containing α -halo carbonyls can provide an alternate approach to the synthesis of β -hydroxy

Received: October 12, 2011 Published: December 2, 2011 Scheme 1. Aldol Reaction Strategies for the Construction of Dap Precursor 8 (eq 1)⁶ and Dil Precursors 10a,b (eq 2)⁹ Employing Chiral Auxiliaries in the Presence (7a and 7c) and Absence (7b) of α -[F](#page-4-0)unctionality

derivatives.¹⁰ However, literature pertaining to the synthesis of $β$ -hydroxy-γ-amino acids by asymmetric Reformatsky reactions employing [o](#page-4-0)xazolidinone chiral uxiliaries is significantly less extensive than for asymmetric aldol reactions. These protocols often involve substituents at the α -position, as exemplified by the synthesis of Dap (4) via a cobalt−phosphine complexdirected Reformatsky reaction.¹¹ Therefore, we noted with interest the ability of $SmI₂$ to mediate Reformatsky reactions of haloacetyloxazolidinones with [ald](#page-5-0)ehydes to form β-hydroxy adducts in high stereoselectivity without the necessity of α substituents (for example, see ref 12). As reported herein, in order to examine the potential utility of this chemistry for the preparation of β-hydroxy-γ-amino [acid](#page-5-0)s lacking α-substituents, we applied SmI₂-catalyzed Reformatsky chemistry to the synthesis of N-Boc-protected isostatine (2b) and Dil (3c).

The synthesis of 2b began with the known aldehyde 11, which was obtained from commercially available N-Boc-D-alloisoleucine.^{2,13} We were pleased to observe that the addition of a mixed solution of aldehyde 11 and α -chloroacetyloxazolidinone (R) -12^{12d} [t](#page-4-0)[o](#page-5-0) an excess (3 equiv) of SmI₂¹⁴ resulted in the formation of the secondary alcohol 13 as a single diastereomer (Sche[me](#page-5-0) 2). Removal of the chiral auxilia[ry](#page-5-0) under oxidative

Scheme 2. Synthesis of N-Boc-isostatine (2b)

conditions¹¹ provided N-Boc-isostatine $(2b)$ as a 3:1 mixture of conformers, as indicated by NMR. The optical rotation of the product a[nd](#page-5-0) the spectral properties of the major conformer agreed well with literature values.

The differentiation of conformers vs stereoisomers was made using NMR. COSY analysis displayed two distinct CH_v−NH correlations and NOESY analysis revealed a same-phase (relative to the diagonal) crosspeak between the two NH signals.¹⁵ This same-phase crosspeak arises from the *conforma*tional exchange of a single diastereomer. The presence of two diaster[eo](#page-5-0)mers would not display such a NH−NH crosspeak.

The synthesis of N-Boc-Dil (3c) started with N-Boc-Nmethylisoleucinal 14. This was obtained from commercially available N -Boc-N-methylisoleucine⁹ in a manner analogous to that used to convert N-Boc-D-allo-isoleucine to N-Boc-Dallo-isoleucinal (13). As noted abo[v](#page-4-0)e, while isostatine and Dil share the same relative β ,*γ-erythro* stereochemistry, they are enantiomeric. Accordingly, α -chloroloacetyloxazolidinone (S)- 12^{12} was used in the Sm-mediated Reformatsky coupling with aldehyde 14. Under similar reaction conditions as above, su[cce](#page-5-0)ssful coupling was again realized to provide the secondary alcohol 15 as a single diastereomer in good yield (Scheme 3).

Subsequent O-methylation using trimethyloxonium tetrafluoro $roborate²$ and oxidative removal of the chiral auxiliary yielded N-Boc-Dil (3c), having spectral and optical properties matching lit[e](#page-4-0)rature values.³

The stereochemical outcome of the Sm^{II}-mediated formation of Reformatsky [pr](#page-4-0)oducts 13 and 15 presumably depends highly on the nature of the chiral auxiliary Sm-enolate complex. However additions of achiral lithium enolates to (S)-prolinal derivatives have also been shown to be highly diastereoselective.¹⁶ To determine whether the selectivities observed with the Sm^{II}-mediated Reformatsky reactions to form products 13 and 1[5](#page-5-0) are derived from the chirality of the Sm^{III} enolate (ie. auxiliary control) or from preferential approach of a Sm^{III} enolate to a low energy conformation of the aldehyde (i.e., Felkin addition),¹⁶ we coupled N-Boc-(S)-prolinal (6) to both α -chloroacetyloxazolidinones (R) -12 and (S) -12 using the above reaction c[on](#page-5-0)ditions (Scheme 4).

Treatment of 6 with oxazolidinones (R) -12 and (S) -12 provided the alcohols 17 and 19, [re](#page-2-0)spectively (Scheme 4). Oxidative removal of the chiral auxiliaries gave the corresponding diastereomeric $β$ -hydroxyacids 18 and 20, respectiv[ely](#page-2-0). Spectral analysis confirmed that the stereochemical outcome of the Sm^{II}-mediated Reformatsky coupling was indeed controlled by the chirality of the auxiliary employed in the reaction, allowing threo $(17; dr 5:1)$ or erythro $(19; dr 14:1)$ products to be obtained selectively. It should be noted that chiral induction by α -amino aldehydes generally favors the *erythro* products.¹⁶ This preference accounts for the difference in selectivity observed in the formation of threo product 17 (dr 5:1) [vs](#page-5-0)

Scheme 4. Stereoselective Synthesis of (R) - and (S) - β -Hydroxy-γ-amino Acids Derived from N-Boc-(S)-prolinal (6)

erythro product 19 (dr 10:1). Utilizing the appropriate chiral auxiliary can enhance this preference (erythro/threo = 10:1 vs 4:1^{16a}) or reverse it (*erythro/threo* = 1:5).

In summary, the SmI₂-mediated Reformatsky coupling of amin[o a](#page-5-0)ldehydes with α -chloroacetyloxazolidinones has been successfully applied to the synthesis of the β -hydroxy- γ -amino acids N-Boc-isostatine (2b) and N-Boc-dolaisoleucine (3c). The stereochemical outcomes of the reactions are controlled by the absolute configuration of the chiral auxiliaries used, allowing the selective formation of erythro or threo products. These results should serve as a general method for the construction of β hydroxy-γ-amino acids that enables the synthesis of both natural and unnatural peptide sequences for biological evaluation.

EXPERIMENTAL SECTION

General Information. Reactions were stirred magnetically under an argon atmosphere, unless otherwise noted, and reagents were purchased from commercial sources and used without further purification. Solvents were removed by rotary evaporation under reduced pressure and silica gel chromatography was performed using flash silica gel (230−400 mesh, 40−60 μm particle size). Anhydrous solvents were obtained commercially and used without further drying. Infared (IR) measurements were made using a Fourier transform infared spectrometer equipped with an ATR probe. NMR measurements were performed at 25 °C (unless otherwise noted) at either 400 or 500 MHz. When required, NMR spectra were acquired at elevated temperatures, specified within the reported data and in the spectral parameters.

N-Boc-p-allo-isoleucinal (11). To a solution of N-Boc-p-alloisoleucine (1 g, 4.32 mmol) in THF (4.32 mL), at −10 °C under argon, was added N-methylmorpholine (0.475 mL, 4.32 mmol), followed by isobutyl chloroformate (0.568 mL, 4.32 mmol). After being stirred at −10 °C (10 min), the reaction mixture was filtered (glass fritted funnel) to remove the precipitate and the filtrate cooled to −10 °C. A solution of NaBH₄ (0.245 g, 6.49 mmol) in H₂O (2.2 mL) was then added over 5 min. Upon complete addition, the reaction was diluted with H₂O (5 mL) and extracted with EtOAc (3 \times 30 mL). The organic phase was dried (Na_2SO_4) and concentrated in vacuo.¹³ The crude residue was diluted with DMSO (4.3 mL) under argon, and NEt_3 (3.01 mL, 21.60 mmol) was added. The mixture was stirred [at](#page-5-0) room temperature (15 min) before being cooled to 0 \degree C. SO₃− pyridine complex (3.438 g, 21.60 mmol) was added, and the mixture was stirred at 0 °C (45 min). The reaction was halted by the addition of H₂O (30 mL). The mixture was extracted with Et₂O (3 \times 60 mL), and the organic phases were washed, successively, with 10% aqueous citric acid, H_2O , saturated aqueous NaHCO₃, and brine (20 mL each).

All aqueous phases were back extracted with $Et₂O$ (1 \times 50 mL). Combined organic fractions were dried (Na_2SO_4) , concentrated in vacuo and chromatographed over silica gel to afford 11 (706 mg, 3.3 mmol, 76% yield) as a colorless oil. Spectroscopic data for compound 11 matched that previously reported.¹

Samarium Iodide (SmI₂). Using standard anhydrous techniques, with careful exclusion of oxygen, to a [ro](#page-5-0)und-bottomed flask was added a dry mixture of samarium metal (99.9% purity, 40 mesh, 3.76 g, 25.00 mmol; 1.25 equiv) and diiodine (5.08 g, 20 mmol) under argon. The flask containing the dry mixture was then quickly evacuated and argon backfilled (3×), anhydrous THF (200 mL) was added via cannula, the flask was wrapped in aluminum foil, and the mixture was stirred at room temperature $(4 h)$. The SmI₂ obtained in this fashion is deep green-blue in color with a nominal concentration of 0.1 M. When protected from light and stored under argon, this stock solution is stable over a period of approximately 1 week without any appreciable loss of activity.

General Procedure (A) for SmI₂-mediated Reformatsky Coupling. To a dry round-bottomed flask, evacuated and backfilled with argon (3×) at -78 °C, was added freshly prepared SmI₂ (3 equiv) followed by a solution of aldehyde (1.2 equiv) and α chloroacetyloxazolidinone (1 equiv) in dry THF (0.3 M), dropwise via syringe. The transfer was quantitated with additional THF, and the mixture was stirred at −78 °C (5 min). The reaction was terminated by bubbling O_2 through the solution to quench residual Sm^{II} (indicated by a change in color from blue-green Sm^H to yellow Sm^{III}). A solution of saturated aqueous NH₄Cl was then added at −78 °C, and the mixture was brought to room temperature and further diluted with aqueous NH4Cl. The mixture was extracted with $Et₂O$, and the combined organic phases were washed thoroughly with 15% aqueous $\text{Na}_2\text{S}_2\text{O}_3$, dried (Na_2SO_4), and concentrated in vacuo. The resulting crude residue was purified by silica gel chromatography to provide the desired coupled products (see below).

(3S,4R,5S)-4-tert-Butyl carbamate-3-hydroxy-1-((R)-4-isopropyl-2-oxazolidinone-3-yl)-5-methyl-1-oxoheptanone (13). The general procedure "A" outlined above was followed using SmI_2 (9.29 mL, 0.929 mmol, 3 equiv), N-Boc-D-allo-isoleucinal (11) (80 mg, 0.372 mmol, 1.2 equiv) and α -chloroacetyloxazolidinone (R)-12 (63.7 mg, 0.310 mmol). The product residue was chromatographed over silica gel to afford alcohol 13 (90.4 mg, 0.234 mmol, 76%) as a colorless oil: TLC $R_f = 0.27$ (40% EtOAc/hexane); α_D^{25} (c 2.5, CHCl₃) = (−) 38.27; ¹H NMR (25 °C, 500 MHz, CDCl₃) δ 4.49− 4.45 (m, 2H), 4.31 (t, $J = 8.6$ Hz, 1H), 4.20 (dd, $J = 9.0$, 2.6 Hz, 1H), 3.95 (bs, 0.1H), 3.86 (ddd, $J = 14.9$, 8.5, 3.3 Hz, 0.9H), 3.69 (td, $J =$ 9.6, 3.5 Hz, 1H), 3.58 (d, J = 8.1 Hz, 1H), 3.34 (dd, J = 17.4, 6.6 Hz, 1H), 3.04 (dd, J = 17.4, 3.3 Hz, 1H), 2.38−2.31 (m, 1H), 1.97 (ddd, J = 13.8, 6.9, 3.6 Hz, 1H), 1.44 (s, 1H), 1.41 (s, 8H), 1.38−1.30 (m, 1H), 1.26−1.19 (m, 1H), 0.92 (d, J = 7.0 Hz, 3H), 0.92−0.87 (m, 3H), 0.88 (d, J = 6.9 Hz, 3H), 0.85 (d, J = 6.9 Hz, 3H); ¹H NMR (75 °C, 500 MHz, DMSO- d_6) δ 4.59 (bs, 1H), 4.41−4.36 (m, 1H), 4.31 (t, $J = 8.6$ Hz, 1H), 4.28–4.27 (m, 1H), 4.25 (dd, $J = 8.9$, 3.0 Hz, 1H), 3.03 (bs, 1H), 2.92 (dd, J = 15.9, 3.4 Hz, 1H), 2.66 (s, 3H), 2.25−2.18 (m, 1H), 1.89−1.81 (m, 1H), 1.50 (bs, 1H), 1.40 (s, 9H), 1.07−1.00 $(m, 1H)$, 0.96 (d, J = 6.8 Hz, 3H), 0.87 (d, J = 7.0 Hz, 3H), 0.85 (t, J = 7.5 Hz, 3H), 0.82 (d, J = 6.9 Hz, 3H); ¹³C NMR (25 °C, 125 MHz, CDCl3) δ 173.6, 156.4, 154.04, 79.5, 69.6, 63.8, 58.6, 56.1, 37.7, 33.7, 28.8, 28.3, 27.1, 18.0, 14.9, 13.1, 11.7; IR (neat film, cm[−]¹) 3371, 2963, 1783, 1697, 1236; HRMS (MALDI - TOF) calcd for $C_{19}H_{34}N_2O_6$ $[M + Na]$ ⁺ 409.2315, found 409.2327.

N-Boc-N-methylisoleucinal (14). To a solution of N-Boc-N-Meisoleucine (2.94 g, 12 mmol) in dry THF (8.4 mL) at 0 $^{\circ}$ C was added BH₃−THF complex (1.0 M in THF, 18 mL, 1.5 equiv) dropwise via syringe. The reaction solution was stirred at $0^{\circ}C(2 h)$ and then room temperature (1 h). The reaction was halted by the slow addition of H₂O (20 mL), and the mixture was extracted with EtOAc (3 \times 50 mL). The organic phase was washed, successively, with saturated aqueous NaHCO₃ (20 mL) and brine (20 mL). Removal of the solvent in vacuo afforded the crude amino alcohol as a viscous clear oil which was diluted with DMSO (10.81 mL) and NEt₃ $(7.53 \text{ mL}, 54.0 \text{ m})$ mmol) under argon. The mixture was stirred at room temperature

(15 min) then cooled to 0 °C. SO₃-pyridine complex (8.60 g, 54.0 mmol) was added and the mixture was stirred at 0 $^{\circ}$ C (45 min). The reaction was halted by the addition of H_2O (50 mL). The mixture was extracted with Et₂O (3×100 mL), and the organic phase was washed, successively, with 10% aqueous citric acid, water, saturated aqueous $NaHCO₃$ and brine (15 mL each). All aqueous phases were backwashed with Et₂O (1 \times 40 mL). Combined organic fractions were dried with Na₂SO₄ and concentrated in vacuo. The residue was eluted through a short plug of silica gel to afford 14 (2.061 g, 8.99 mmol, 75% yield) as a colorless oil. Spectroscopic data for compound 14 matched that previously reported.⁶

(3S,4R,5S)-4-((tert-Butoxycarbonyl)amino)-3-hydroxy-5 methylheptanoic Acid (N-Boc-is[os](#page-4-0)tatine, 2b). To a solution of amide 13 (50 mg, 0.129 mmol) in 3:1 THF/H₂O (0.35 mL) at 0 °C was added 50% aqueous H_2O_2 (0.056 mL, 0.776 mmol) followed by LiOH−H₂O (14.13 mg, 0.336 mmol). The mixture was stirred at 0 $^{\circ}$ C (3 h), and then excess peroxide was quenched by the addition of 1.5 N aqueous Na_2SO_3 (1 mL, 1.5 mmol) at 0 °C, and the mixture was stirred at room temperature (overnight). The pH was adjusted to ∼9− 10 by the addition of saturated aqueous $NaHCO₃$ and the free oxazolidinone side product was extracted using DCM. The aqueous phase was acidified to pH ∼2 using 1 N aqueous HCl and extracted with EtOAc . The combined EtOAc phases were dried (Na_2SO_4) and taken to dryness in vacuo to afford N-Boc-isostatine (2b) (30.3 mg, 0.11 mmol, 85%) as a colorless semisolid: α_{D}^{25} (c 1.25, CHCl₃) = (-) 7.5; ¹H NMR (25 °C, 500 MHz, CDCl₃, 2 conformers) δ 5.77 (NH, d, $J = 10.6$ Hz, 0.36H, minor conformer, exchanges), 4.51 (NH, d, $J = 9.6$) Hz, 0.64H, major conformer, exchanges), 3.95 (td, $J = 8.1$, 3.0 Hz, 1H), 3.65 (td, J = 9.7, 3.8 Hz, 0.66H, major), 3.58 (t, J = 8.6 Hz, 0.33H, minor), 2.70−2.62 (m, 1H), 2.57−2.47 (m, 1H), 1.92−1.85 (m, 1H), 1.47 (s, 3H, minor), 1.44 (s, 6H, major), 1.40−1.32 (m, 1H), 1.26−1.19 (m, 2H), 0.92 (t, J = 7.4 Hz, 3H), 0.89−0.85 (m, 3H); ¹³C NMR (25 °C, 125 MHz, CDCl₃) δ major 176.65, 156.6, 80.0, 69.1, 56.8, 38.3, 33.9, 28.31, 27.0, 13.3, 11.6; minor 176.67, 157.8, 81.4, 68.5, 58.0, 39.2, 34.1, 28.26, 26.8, 12.8, 11.8; LRMS (ESI−negative mode) m/z 274 [M – H]⁻, 310 [M + ³⁵Cl]⁻, 312 [M + ³⁷Cl]⁻; LRMS (ESI– positive mode) m/z 298 $[M + Na]$ ⁺; HRMS (MALDI - TOF) calcd for $C_{13}H_{25}NO_5$ [M + Na]⁺ 298.1630, found 298.1640.

(3R,4S,5S)-4-(tert-Butoxycarbonyl)(methyl)amino-3-hydroxy-1-((S)-4-isopropyl-2-oxazolidinone-3-yl)-5-methyl-1-oxoheptanone (15). The general procedure "A" outlined above was followed using SmI_2 (136 mL, 13.60 mmol, 3 equiv), N-Boc-N-Meisoleucinal 14 (1.247 g, 5.44 mmol, 1.2 equiv), and α -chloroacetyloxazolidinone (S)-12 (0.932 g, 4.53 mmol). The product residue was chromatographed over silica gel to afford alcohol 15 (1.54 g, 3.85 mmol, 86%) as a colorless oil: TLC $R_f = 0.22$ (35% EtOAc/hexane); $\alpha_{\rm D}^{\rm 25}$ (c 2.6, CHCl₃) = (−) 38.71; ¹H NMR (25 °C, 500 MHz, DMSO d_6) δ 4.84 (dd, J = 7.3, 28.7 Hz, 1H), 4.40–4.35 (m, 1H), 4.32–4.20 (m, 3H), 3.76 (bs, 1H), 2.99−2.85 (m, 1H), 2.91 (d, J = 6.2 Hz, 1H), 2.64 (s, 1.6H), 2.59 (s, 1.3H), 2.22−2.14 (m, 1H), 1.82 (m, 1H), 1.53−1.43 (m, 1H), 1.38 (s, 9H), 1.04−0.95 (m, 1H), 0.94−0.90 (m, 3H), 0.87–0.78 (m, 9H); ¹H NMR (75 °C, 500 MHz, DMSO- d_6) δ 4.61 (bs, 1H), 4.41–4.37 (m, 1H), 4.31 (t, J = 8.6 Hz, 1H), 4.29–4.23 (m, 1H), 4.25 (dd, J = 8.9, 2.8 Hz, 1H), 3.81−3.59 (m, 1H), 3.01 (bs, 1H), 2.92 (dd, J = 15.7, 3.1 Hz, 1H), 2.66 (s, 3H), 2.22 (dtd, J = 13.8, 6.9, 4.0 Hz, 1H), 1.86 (dd, J = 13.4, 6.7 Hz, 1H), 1.55−1.45 (m, 1H), 1.40 (s, 9H), 1.08–0.98 (m, 1H), 0.96 (d, J = 6.8 Hz, 3H), 0.87 (d, J = 7.0 Hz, 3H), 0.88−0.84 (m, 3H), 0.82 (d, J = 6.9 Hz, 3H); 13C NMR (75 °C, 125 MHz, d_6 -DMSO) δ 170.7, 153.5, 78.1, 67.1, 63.3, 57.8, 40.6, 34.6, 34.2, 28.1, 27.7, 25.0, 17.1, 16.0, 14.5, 10.9; IR (neat film, cm[−]¹) 3458, 2964, 1780, 1684; HRMS (MALDI - TOF) calcd for $C_{20}H_{36}N_2O_6$ [M + Na]⁺ 423.2471, found 423.2477.

 $\widetilde{A}(3R.A.S.S)$ -4-tert-Butoxycarbonyl)(methyl)amino-1-((S)-4isopropyl-2-oxazolidinone-3-yl)-3-methoxy-5-methyl-1-oxoheptanone (16). A mixture of alcohol 15 (304 mg, 0.76 mmol) and molecular sieves (4 Å, oven-dried, 300 mg) was diluted with anhydrous 1,2-dichloroethane (1.12 mL) under an argon atmosphere and stirred (20 min). The mixture was cooled to 0° C prior to the sequential addition of proton sponge [1,8-bis(dimethylamino) naphthalene; 423 mg, 1.97 mmol, 2.6 equiv] and trimethyloxonium

tetrafluoroborate (281 mg, 1.90 mmol, 2.5 equiv). The mixture was brought to room temperature, sealed under argon, and stirred (48 h). The mixture was filtered through Celite and concentrated in vacuo, and the residue was chromatographed over silica gel to afford methyl ether 16 (271 mg, 0.66 mmol, 86%) as a colorless oil: TLC $R_f = 0.42$ (35% EtOAc/hexane); α_{D}^{20} (c 0.9, CHCl₃) = (+) 36.0; ¹H NMR (25 °C, 400 MHz, DMSO- d_6 , two conformers) δ 4.41–4.36 (m, 1H), 4.34−4.26 (m, 2H), 4.03−3.85 (m, 2H), 3.26 (s, 1.6H), 3.23 (s, 1.4H), 3.08−3.02 (m, 2H), 2.62 (s, 1.7H), 2.60 (s, 1.3H), 2.22−2.13 (m, 1H), 1.84−1.71 (m, 1H), 1.38 (s, 9H), 1.41−1.34 (m, 1H), 1.08−0.95 (m, 1H), 0.89 (d, J = 6.6 Hz, 3H), 0.87-0.82 (m, 6H), 0.79 (m, 3H); ¹H NMR (60 °C, 400 MHz, DMSO) δ 4.42–4.37 (m, 1H), 4.33 (t, J = 8.5 Hz, 1H), 4.27 (dd, J = 8.9, 2.9 Hz, 1H), 4.00−3.93 (m, 1.4H), 3.85 (bs, 0.4H), 3.26 (s, 3H), 3.14−3.00 (m, 2H), 2.64 (s, 3H), 2.26−2.15 (m, 1H), 1.84−1.73 (m, 1H), 1.45−1.38 (m, 1H), 1.39 (s, 9H), 1.04 $(m, 1H)$, 0.92 $(d, J = 6.7 \text{ Hz}, 3H)$, 0.87 $(d, J = 7.0 \text{ Hz}, 3H)$, 0.86–0.82 (m, 3H), 0.81 (d, J = 6.9 Hz, 3H); ¹³C NMR (60 °C, 100 MHz, DMSO- d_6 , major conformer) δ 170.4, 153.7, 76.9, 63.3, 57.9, 37.3, 33.2, 27.8, 25.3, 17.2, 15.8, 14.5, 10.4; IR (neat film, cm[−]¹) 2965, 1777, 1688, 1150; HRMS (MALDI - TOF) calcd for $C_{21}H_{38}N_{2}O_{6}$ [M + Na]⁺ 437.2628, found 437.2632.

(3R,4S,5S)-4-((tert-Butoxycarbonyl)(methyl)amino)-3-methoxy-5-methylheptanoic Acid (N-Boc-Dolaisoleucine, 3c). To a solution of carbamate 16 (261 mg, 0.548 mmol) in a 1:1 mixutre of THF/H₂O (1.26 mL), under argon and at 0 $^{\circ}$ C, was added 50% aqueous H_2O_2 (0.114 mL, 1.972 mmol) over 5 min. The resultant solution was stirred (5 min), LiOH−H2O (36.8 mg, 0.876 mmol) was added, and stirring was continued at 0 $^{\circ}$ C (3 h). The reaction was quenched by the addition of Na_2SO_3 (276 mg, 2.191 mmol, 4 equiv) in H_2O (1.5 mL, 1.46 M), and the mixture was stirred at room temperature (overnight). The mixture was concentrated in vacuo, and the resulting aqueous slurry was washed with DCM. The aqueous phase was acidified to pH ∼2 using 37% aqueous HCl and extracted with EtOAc. The original DCM extracts were again extracted with 5% NaOH (10 mL), and the aqueous layer was acidified and extracted (EtOAc) as before. The DCM organic phases were dried (Na_2SO_4) and concentrated in vacuo to afford recovered chiral auxiliary (64 mg, 90%). The combined EtOAc organic phases were dried (Na_2SO_4) and taken to dryness in vacuo to afford 3c (158 mg, 0.52 mmol, 95%) as a viscous oil: α_{D}^{19} (c 2.5, CHCl₃) = (-) 10.9; ¹H NMR (25 °C, 400 MHz, CDCl₃) δ 4.02−3.80 (m, 2H), 3.38 (s, 3H), 2.67 (s, 3H), 2.60− 2.43 (m, 2H), 1.82−1.69 (m, 1H), 1.51−1.40 (m, 1H), 1.43 (s, 9H), 1.12−1.02 (m, 1H), 0.94 (d, J = 5.9 Hz, 3H), 0.87 (t, J = 7.3 Hz, 3H); ¹³C NMR (25 °C, 100 MHz, CDCl₃) δ 176.5 and 176.4, 156.6, 80.1 and 79.6, 78.3, 60.9 (br), 57.7 and 57.6, 37.1 and 36.9, 34.9 (br) and 34.5, 28.39 and 28.35, 25.9 and 25.7, 16.2 and 16.1, 11.28; HRMS (MALDI - TOF) calcd for $C_{15}H_{29}NO_5$ [M + Na]⁺ 326.1943, found 326.1916.

(S)-tert-Butyl 2-((S)-1-Hydroxy-3-((R)-4-isopropyl-2-oxoxazolidin-3-yl)-3-oxopropyl)pyrrolidine-1-carboxylate (17) and (S) tert-Butyl 2-((R)-1-Hydroxy-3-((R)-4-isopropyl-2-oxoxazolidin-3-yl)-3-oxopropyl)pyrrolidine-1-carboxylate (epi-17). The general procedure "A" outlined above was followed using SmI_2 (29.2 mL, 2.92 mmol, 3.0 equiv), N-Boc-prolinal 6 (233 mg, 1.168 mmol, 1.2 equiv), and (R) -12 (200 mg, 0.973 mmol, 1.0 equiv). The product residue was chromatographed over silica gel to afford a mixture (310 mg, 0.84 mmol, 86%, dr 5:1) of alcohols 17 and *epi-*17: TLC $R_f = 0.34$ $(55\% \text{ EtOAc/hexane})$; α_{D}^{23} (c 1.4, CHCl₃) = (-) 94.5; 17 ¹H NMR (25 °C, 400 MHz, DMSO- d_6) δ 4.90 (s, 0.1H), 4.81 (d, J = 5.9 Hz, 0.9H), 4.39−4.32 (m, 1H), 4.33−4.22 (m, 2.3H), 4.12 (bs, 0.6H), 3.95−3.75 (m, 0.8H), 3.61 (bs, 0.1H), 3.37−3.28 (m, 1H), 3.26−2.99 (m, 1.7H), 2.99−2.60 (m, 1.3H), 2.27−2.12 (m, 1H), 1.94−1.77 (m, 2.8H), 1.75−1.63 (m, 1.2H), 1.40 (s, 1.7H), 1.38 (s, 7.3H), 0.85 (d, J = 7.0 Hz, 3H), 0.82−0.77 (m, 3H); ¹ H NMR (60 °C, 400 MHz, DMSO- d_6) δ 4.68 (d, J = 5.2 Hz, 0.86H), 4.39–4.35 (m, 1H), 4.30 (t, J = 8.4 Hz, 1H), 4.25 (dd, J = 8.9, 3.1 Hz, 1H), 4.25−4.17 (m, 1H), 3.88 (bs, 0.86H), 3.42−3.33 (m, 1H), 3.24−3.14 (m, 1.4H), 3.04− 2.82 (m, 1.6H), 2.26−2.17 (m, 1H), 1.90−1.81 (m, 3H), 1.75−1.68 $(m, 1H)$, 1.40 $(s, 7.7H)$, 0.86 $(d, J = 7.1 \text{ Hz}, 3H)$, 0.83 $(d, J = 6.8 \text{ Hz},$ 3H); ¹³C NMR (60 °C, 100 MHz, DMSO- d_6) δ 171.4, 154.3, 78.8,

63.8, 58.4, 47.5, 28.8, 28.5, 27.2, 17.9, 15.2; IR (neat film, cm[−]¹) 2972, 2360, 1780, 1685; HRMS (MALDI - TOF) calcd for $C_{18}H_{30}N_2O_6$ $[M + K]^+$ 409.1741, found 409.1719. epi-17: diagnostic ¹H NMR peaks (60 °C, 400 MHz, DMSO- d_6) δ 4.73 (d, J = 5.7 Hz, 0.14H), 3.65 (bs, 0.14H), 2.73 (dd, $J = 15.8$, 3.3 Hz, 0.2H), 1.41 (s, 1.3H).

(S)-3-((S)-1-(tert-Butoxycarbonyl)pyrrolidin-2-yl)-3-hydroxypropanoic Acid (18). To a solution of carbamate 17 (70 mg, 0.189 mmol) in 4:1 THF/H₂O (1.08 mL, 0.175 M), at 0 °C, was added 50% aqueous H_2O_2 (39 μ L, 0.680 mmol, 3.6 equiv) dropwise via syringe. A solution of LiOH−H₂O (12.70 mg, 0.302 mmol, 1.6 equiv) in H₂O (216 μ L) was added, and stirring was continued at 0 °C (3 h). A solution of Na₂SO₃ (95 mg, 0.756 mmol, 4 equiv) in H₂O (518 μ L, 1.46 M) was added, and the mixture was stirred at room temperature (overnight). The mixture was concentrated in vacuo, and the residue was diluted with 5% aqueous NaHCO₃ (10 mL). The aqueous phase was washed with DCM (2 × 10 mL, acidified to pH ∼2 with 37% aqueous HCl and extracted with EtOAc. The original DCM washes were extracted with 5% aqueous NaHCO₃ (10 mL), and the aqueous layer was acidified and extracted with EtOAc. The DCM phases were dried (Na_2SO_4) and concentrated in vacuo to afford recovered chiral auxiliary (23 mg, 0.178 mmol, 94% recovered). The combined EtOAc phases were dried (Na_2SO_4) and taken to dryness in vacuo to afford a mixture (46.4 mg, 0.179 mmol, 95%, $dr = 5:1$) of acids 18 and 20 as a colorless oil: α_{D}^{20} (c 2.3, MeOH) = (-) 47.1; 18⁻¹H NMR (25 °C, 400 MHz, DMSO-d6) δ 4.27−4.17 (m, 0.45H), 4.17−4.07 (m, 0.55H), 3.85−3.75 (m, 0.84H), 3.39−3.25 (m, 1H), 3.24−3.11 (m, 1H), 2.32−2.19 (m, 1H), 2.13 (dd, J = 15.0, 9.3 Hz, 1H), 1.91−1.73 (m, 3H), 1.73−1.63 (m, 1H), 1.39 (s, 9H); ¹ H NMR (65 °C, 400 MHz, DMSO- d_6) δ 4.23–4.08 (m, 1H), 3.86–3.79 (m, 0.83H), 3.41– 3.31 (m, 1H), 3.22−3.12 (m, 1H), 2.28 (dd, J = 15.0, 4.0 Hz, 1H), 2.16 (dd, J = 15.1, 9.0 Hz, 1H), 1.89−1.76 (m, 3H), 1.74−1.64 (m, 1H), 1.41 (s, 9H); ¹³C NMR (65 °C, 100 MHz, DMSO-d₆) δ 172.6, 154.2, 78.3, 68.9, 60.3, 46.8, 37.9, 27.9, 26.2, 23.2; IR (neat film, cm⁻¹) 3402, 2930, 2361, 1677, 1394; HRMS (MALDI - TOF) calcd for $C_{12}H_{21}NO_5 [M + Na]^+$ 282.1317, found 282.1318. 20: diagnostic ¹H NMR peak (25 °C, 400 MHz, DMSO-d₆) δ 3.62-3.56 (m, 0.16H); diagnostic ¹H NMR peak (65 °C, 400 MHz, DMSO- d_6) δ 3.66–3.60 $(m. 0.16H)$.

(S)-tert-Butyl 2-((R)-1-Hydroxy-3-((S)-4-isopropyl-2-oxooxazolidin-3-yl)-3-oxopropyl)pyrrolidine-1-carboxylate (19) and (S)-tert-Butyl 2-((S)-1-Hydroxy-3-((S)-4-isopropyl-2-oxooxazolidin-3-yl)-3-oxopropyl)pyrrolidine-1-carboxylate (epi-19). The general procedure "A" outlined above was followed using SmI2 (29.200 mL, 2.92 mmol), N-Boc-prolinal 6 (233 mg, 1.168 mmol), and (S)-12 (200 mg, 0.973 mmol). The product residue was chromatographed over silica gel to afford pure alcohol 19 (272 mg, 0.75 mmol, 74%). An additional amount (71 mg) of 19 and epi-19 was also obtained as a mixture (ratio 19:*epi*-19 = 53:47). 19: TLC R_f = 0.35 (55% EtOAc/hexane); $\alpha_{\rm D}^2$ (c 0.79, CHCl₃) = (+) 17.1; ¹H NMR (25 °C, 400 MHz, DMSO-d₆) δ 4.94−4.83 (m, 1H), 4.42−4.33 (m, 1H), 4.33−4.24 (m, 2H), 4.24−4.14 (m, 1H), 3.69−3.57 (m, 1H), 3.34−3.27 (m, 1H), 3.22−3.08 (m, 1H), 3.07−2.77 (m, 2H), 2.24− 2.12 (m, 1H), 1.99−1.80 (m, 2H), 1.78−1.64 (m, 2H), 1.40 (s, 9H), 0.85 (d, J = 7.0 Hz, 3H), 0.79 (d, J = 6.9 Hz, 3H); ¹H NMR (60 °C, 400 MHz, DMSO- d_6) δ 4.75 (d, J = 5.0 Hz, 1H), 4.41–4.36 (m, 1H), 4.31 (t, J = 8.5 Hz, 1H), 4.26 (dd, J = 9.0, 3.1 Hz, 1H), 4.23−4.18 (m, 1H), 3.70−3.62 (m, 1H), 3.41−3.31 (m, 1H), 3.20−3.12 (m, 1.2H), 3.04−2.88 (m, 1.8H), 2.25−2.15 (m, 1H), 1.99−1.90 (m, 1H), 1.90− 1.81 (m, 1H), 1.80−1.65 (m, 2H), 1.41 (s, 9H), 0.86 (d, J = 7.0 Hz, 3H), 0.81 (d, J = 6.9 Hz, 3H); ¹³C NMR (60 °C, 100 MHz, DMSOd6) δ 170.5, 153.6, 78.1, 67.5, 63.2, 61.1, 57.7, 46.4, 28.2, 27.93, 25.2, 17.20, 14.56; IR (neat film, cm[−]¹) 3373, 2969, 2360, 1781, 1685; HRMS (MALDI - TOF) calcd for $C_{18}H_{30}N_2O_6$ [M + Na]⁺ 393.2002, found 393.1994. epi -19: diagnostic $^1\mathrm{H}$ NMR peaks (60 $^\circ\mathrm{C}$, 400 MHz, DMSO- d_6) δ 4.68 (d, J = 4.7 Hz), 3.86 (d, J = 3.5 Hz), 1.40 (s), 0.82 (d, J = 6.9 Hz); diagnostic ¹³C NMR peaks (100 MHz, DMSO- d_6) δ 170.9, 153.8, 78.2, 57.9, 46.9, 28.3, 27.88, 26.5, 17.22, 14.61.

(R)-3-((S)-1-(tert-Butoxycarbonyl)pyrrolidin-2-yl)-3-hydroxypropanoic Acid (20). To a solution of carbamate 19 (70 mg, 0.189 mmol) in 4:1 THF:H₂O (1.08 mL, 0.175 M), at 0 °C, was added 50% aqueous H_2O_2 (39 μ L, 0.680 mmol, 3.6 equiv) dropwise via syringe. A solution of LiOH-H₂O (12.70 mg, 0.302 mmol, 1.6 equiv) in H_2O (216 μ L) was added and the reaction solution was stirred at 0 °C (3 h). To this solution was added a solution of Na_2SO_3 (95 mg, 0.756) mmol, 4 equiv) in H₂O (518 μ L, 1.46 M) and the mixture was stirred at room temperature (overnight). The mixture was concentrated in *vacuo*, and the residue was diluted with 5% aqueous NaHCO₃ (10 mL) and washed with DCM $(2 \times 10 \text{ mL})$. The resulting aqueous layer was acidified to pH∼2 using 37% aqueous HCl and extracted with EtOAc. The original DCM washes were extracted with 5% aqueous $NAHCO₃$ (10 mL), and the resultant aqueous layer was acidified and extracted with EtOAc. The DCM phases were dried (Na_2SO_4) and concentrated in vacuo to afford recovered chiral auxiliary (23 mg, 0.178 mmol, 94% recovered). The combined EtOAc phases were dried $(Na₂SO₄)$ and taken to dryness in vacuo to afford acid 20 (46.8 mg, 0.18 mmol, 96%) as a pale red oil. $\alpha_{\rm D}^{20}$ (c 2.3, CHCl₃) = (−) 46.3; ¹H NMR (65 °C, 400 MHz, d_6 -DMSO) δ 4.14−4.06 (m, 1H), 3.66−3.60 (m, 1H), 3.38−3.29 (m, 1H), 3.20−3.10 (m, 1H), 2.28 (dd, J = 14.9, 4.2 Hz, 1H), 2.18 (dd, J = 14.8, 8.7 Hz, 1H), 1.97−1.81 (m, 2H), 1.77− 1.64 (m, 2H), 1.41 (s, 9H); ¹³C NMR (65 °C, 100 MHz, d_6 -DMSO) δ 172.8, 153.7, 78.0, 67.8, 61.0, 61.0, 46.4, 39.8, 28.0, 25.0, 23.2; IR (neat film, cm[−]¹) 3397, 2972, 1676, 1401; HRMS (MALDI - TOF) calcd for $C_{12}H_{21}NO_5 [M + Na]^+$: 282.1317, found 282.1318.

■ ASSOCIATED CONTENT

6 Supporting Information

 1 H and 13 C NMR spectra for all compounds are provided. This material is available free of charge via the Internet at http:// pubs.acs.org.

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■ REFERENCES

(1) Umezawa, H.; Aoyagi, T.; Morishima, H.; Matsuzaki, M.; Hamada, M.; Takeuchi, T. J. Antibiot. 1970, 23, 259.

(2) Rinehart, K. L.; Sakai, R.; Kishore, V.; Sullins, D. W.; Li, K. M. J. Org. Chem. 1992, 57, 3007.

(3) Shioiri, T.; Hayashi, K.; Hamada, Y. Tetrahedron 1993, 49, 1913. (4) Boger, D. L.; Colletti, S. L.; Honda, T.; Menezes, R. F. J. Am. Chem. Soc. 1994, 116, 5607.

(5) Ordonez, M.; Cativiela, C. Tetrahedron: Asymmetry 2007, 18, 3. (6) Pettit, G. R.; Burkett, D. D.; Barkoczy, J.; Breneman, G. L.; Pettit, W. E. Synthesis 1996, 719.

(7) Owa, T.; Haupt, A.; Otsuka, M.; Kobayashi, S.; Tomioka, N.; Itai, A.; Ohno, M.; Shiraki, T.; Uesugi, M.; Sugiura, Y.; Maeda, K. Tetrahedron 1992, 48, 1193.

(8) Kolodiazhnyi, O. I. Tetrahedron 2003, 59, 5953.

(9) Pettit, G. R.; Burkett, D. D.; Williams, M. D. J. Chem. Soc., Perkin Trans. 1 1996, 853.

(10) Ocampo, R.; Dolbier, W. R. Jr. Tetrahedron 2004, 60, 9325.

(11) Pettit, G. R.; Grealish, M. P. J. Org. Chem. 2001, 66, 8640.

(12) (a) Orsini, F.; Sello, G.; Manzo, A. M.; Lucci, E. M. Tetrahedron: Asymmetry 2005, 16, 1913. (b) Kim, S.-J.; Kang, H.-Y.; Sherman, D. H. Synthesis 2001, 2001, 1790. (c) John, J. P.; Jost, J.; Novikov, A. V. J. Org. Chem. 2009, 74, 6083.

(13) Hili, R.; Rai, V.; Yudin, A. K. J. Am. Chem. Soc. 2010, 132, 2889.

 (14) While SmI₂ is commercially available, freshly prepared material is ciritical to obtain reproducible results. A full experimental description is provided in the Experimental Section, which largely follows the method described by: Beemelmanns, C.; Reissig, H.-U. Angew. Chem., Int. Ed. Engl. 2010, 49, 8021.

(15) COSY and NOESY NMR spectra of compound 2b are included in the Supporting Information. (a) Lloyd-Williams, P.; Monerris, P.; Gonzalez, I.; Jou, G.; Giralt, E. J. Chem. Soc., Perkin Trans. 1 1994, 1969. While not reported directly, identical confomeric NMR signals are ob[served within the Supplem](#page-4-0)entory Information, the ¹H spectrum for compound 2b as reported by: (b) Fuse, S.; Okada, K.; Iijima, Y.; Munakata, A.; Machida[, K.; Takahashi, T.; Takagi,](#page-4-0) M.; Shin-ya, K.; Doi, T. Org. Biomol. Chem. 2011, 9, 3825.

(16) (a) Hanson, G. J.; Baran, J. S.; Lindberg, T. Tetrahedron Lett. 1986, 27, 3577. (b) Snider, B. B.; Gao, X. Org. Lett. 2005, 7, 4419.

(17) Rinehart, K. L.; Sakai, R.; Kishore, V.; Sullins, D. W.; Li, K. M. J. Org. Chem. 1992, 57, 3007.